

Original Research Article

Apolipoprotein-E Gene Polymorphism and Possible role of *ApoE* ϵ 4 Allele with a Lower Probability of Progression to HCV-Related Liver Cirrhosis in Egyptian Patients

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Abstract

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Apolipoprotein E (*ApoE*) plays an important role in regulating lipid and lipoprotein metabolism and *ApoE* genotypes are known to affect lipoprotein concentrations. *ApoE* may become a major variable of preventive and personalized medicine due to several biological role. *ApoE* genotypes could be an important host genetic factor affecting disease progression in chronic liver disease. We investigated whether *ApoE* gene polymorphism determines the disease progression to liver cirrhosis in hepatitis C virus (HCV)-infected Egyptian individuals. This case-controlled study enrolled 120 subjects, 80 chronic hepatitis C (CHC) related liver disease patients and 40 age and sex matched healthy control subjects. *ApoE* genotypes were determined by Restriction Fragment Length Polymorphism (RFLP). Restriction isotyping using restriction enzyme (HhaI). Among the 120 subjects, the most common genotype was ϵ 3/ ϵ 3, accounting for (91.67)%, followed by ϵ 3/ ϵ 4 (8.33) %. The genotypes of ϵ 2/2 ϵ 3/3 ϵ 4/4 were not detected in our results. The ϵ 3 allele was the most common allele overrepresented in CHC LC/LF (98.75) % versus (93.75) % in non cirrhotic group. However, CHC non cirrhotic patients had a higher *ApoE* ϵ 4 allele frequency (6.25) % than those with severe disease (1.25) %. Major contribution of *ApoE* ϵ 4 allele was with decreased susceptibility to LC/LF development cannot be ruled out; (OR) (CI95), 0.18(0.02 to 1.61)/0.19 (0.02 to 1.66) (p=0.12,0.13) for (ϵ 3/ ϵ 4) genotype and ϵ 4 allele frequency respectively. Although non statistically significant difference due to relatively small sample size. Our result support other studies for a possible genetic association between *ApoE* ϵ 4 allele with a lower probability of progression to HCV-related liver cirrhosis.

Keywords: Apolipoprotein E, CHC, Cirrhosis, Genotype, HCV, Liver failure

Abbreviation: Apolipoprotein E (**ApoE**); Chronic hepatitis C virus (**CHC**); Hepatitis C Virus (**HCV**); Hepatocellular Carcinoma (**HCC**); Liver Cirrhosis (**LC**); Liver Failure (**LF**); Polymerase Chain Reaction (**PCR**); Restriction Fragment Length Polymorphism (**RFLP**); Single Nucleotide Polymorphism (**SNP**)

INTRODUCTION

Hepatitis C continues to be a major public health problem, during chronic hepatitis C, progressive fibrosis

deposition occurs and this deposition ends in cirrhosis in 20-30% of Chronic Hepatitis C (CHC) carriers and 2.5%

of HCV-infected individuals develop Hepatocellular Carcinoma (HCC) later in life (Poynard et al., 1997; Bowen and Walker, 2005). The pathway of Hepatitis C Virus (HCV) assembly and secretion is closely linked to lipoprotein production and secretion, and HCV particles circulating in the blood associate with lipoproteins, thus, it is termed a Lipo-Viro Particle (LVP) (Nielsen et al., 2006; Hishiki et al., 2010; Aizawa et al., 2015). Moreover, HCV might gain entry into cells via a hitchhiker method with the lipoproteins (Agnello et al., 1999). Specifically, entry might involve LDL receptors (LDLRs) (Aizawa et al., 2015). The key lipoprotein molecules mediating these interactions are apolipoproteins (Sabile et al., 1999; Aizawa et al., 2015). Apolipoprotein E is a key molecule required for HCV entry and is one of the possible therapeutic targets for interrupting HCV infection. Apolipoproteins (Apo) are amphipathic protein on the surface of a lipoprotein particle, which help stabilize lipoprotein structure with numerous roles in regulating lipid and lipoproteins, apolipoproteins are classified into A, B, C, E (Aizawa et al., 2015). *ApoE* is polymorphic proteins, identified in 1973 (Shore and Shore, 1973) and was first recognized for its role in cardio-vascular diseases (CVD) and widely searched not only for cardio-vascular disorders but also in relation to several other medical conditions such as neurodegenerative and autoimmune diseases. The key protein, playing many roles in lipid absorption, transport, local homeostasis in the vessel walls, and endothelial function (Sacre et al., 2003). *ApoE* plays a central role in cholesterol transport as reflected by the association of *ApoE* with a variety of lipoprotein size classes and the ability of *ApoE* to interact with two distinct hepatic receptors (LDL receptor and ApoE receptor) (Ahn et al., 2012; Aizawa et al., 2015). "*ApoE* has additional roles, as a modulator of the immune function (Kelly et al., 1994) and also has direct effect on tissue macrophage recruitment, independently, of the lipoprotein metabolism. Several data on the role of *ApoE* in the regulation of inflammation were reported" (Aizawa et al., 2015).

ApoE is synthesized primarily by the liver and it is estimated that 20-40% of total *ApoE* is produced by extrahepatic tissues with the brain glial cells and macrophages expressing relatively high amounts, with lesser amounts produced by the kidneys, adrenals, spleen, testis and the skin (Eichner et al., 2002; Minihane et al., 2007). The human *ApoE* gene spans 3.7 kb including four exons and three introns (Das et al., 1985; Hixson and Vernier, 1990) and is located on chromosome 19 in a gene family that also contains the genes for apoC-I, C-I' (a pseudo-gene), and C-II (Hixson and Vernier, 1990; Myklebost and Rogne, 1988). *ApoE* exists in 3 major isoforms (ApoE2, ApoE3, ApoE4), determined by two single nucleotide polymorphisms (SNPs) in the *APOE* gene (rs7412 and rs429358, coded by three codominant alleles (ϵ 2, ϵ 3, ϵ 4). The three major alleles are responsible for three homozygous (ϵ 2/ ϵ 2, ϵ 3/ ϵ 3,

4/ ϵ 4), and three heterozygous (ϵ 2/ ϵ 3, ϵ 2/ ϵ 4, ϵ 3/ ϵ 4) genotypes (Weisgraber et al., 1981; Zannis and Breslow, 1981). *ApoE* ϵ 3 seems to be the wild-type isoform with normal function. *ApoE* ϵ 2 and *ApoE* ϵ 4 differ from *ApoE* ϵ 3 by a single amino acid substitution. While *ApoE* ϵ 3 contains cysteine at amino acid position 112 and arginine at position amino acid 158, the arginine is substituted by a cysteine in *ApoE* ϵ 2 carriers and the cysteine is substituted by an arginine in *ApoE* ϵ 4 carriers. Additionally, two minor alleles of the gene, ϵ 1 and ϵ 5, exist but these are present in less than 0.1% of the population (Ordovas et al., 1987). Each isoform has specific functional properties including different susceptibilities to diseases (Myklebost and Rogne, 1988; Bennet et al., 2007; Villeneuve et al., 2014). The reasons for the diversity of outcomes of HCV infection are unclear. Different factors may affect the outcome of HCV (e.g. age, other coexisting virus infections, genetic predisposition etc).

In this study, we evaluate association of the different genotypes of *ApoE* and HCV induced liver cirrhosis in sample size of chronic hepatitis Egyptian patients.

SUBJECTS AND METHODS

Ethical Statement

Informed written consent is obtained from all participants according to human ethics committee approval. The study protocol was reviewed and approved by the Ethics Committee of the Ain Shams University and the work done at Medical Research Center, Ain-Shams University Hospitals.

Subjects

One hundred and twenty subjects were included in our study and were categorized into three groups. The study was done in the period between June 2011 and February 2013. It included 80 Egyptian patients who were recruited from Internal Medicine and Hepatology outpatient clinics and inpatient at Ain Shams University Hospitals, in addition to a control group of 40 healthy volunteers.

Patient groups

Group I: Forty CHC with no signs of cirrhosis.

Group II: Forty CHC cirrhotic patients with signs of liver cell failure or portal hypertension according the modified Child-pugh score (C).

Control group

Forty age and sex matched healthy volunteers to deter-

Table 1. Patients characteristics of the study groups

	Control n=40 n(%)	CHC (non cirrhotic) n=40 n(%)	CHC (Cirrhotic/Liver Failure) n=40 n(%)	Chi-square	
				X ²	P-value
Sex					
Male	20(50)	24(60%)	21(52.5)	0.873	0.646
Female	20(50)	16(40)	19(47.5)		
Liver/US					
Cirrhosis	0	0	40 (100)	120	<0.001
HFL	0	0	0(0)		
No change	40 (100)	40 (100)	0		
Spleen/US					
Normal size	40 (100)	40 (100)	0	120	<0.001
Splenomegaly	0	0	40 (100)		
Encephalopathy					
No attack	40 (100)	40 (100%)	22 (55)	42.353	<0.001
Previous attack	0	0	18 (45)		
Ascites					
No/Ascites	40 (100)	40(100)	0	120	<0.001
Mild	0	0(100)	5 (12.50)		
Moderate	0	0	10 (25)		
Tense/Severe	0	0	25 (62.5)		
Child Pugh Score					
A			0	-	-
B			0		
C			40 (100)		
<i>HFL Hepatic focal lesion Significance < 0.05</i>					

mine the prevalence of *ApoE* different genotypes in Egyptian population.

Exclusion criteria

Patients with liver cirrhosis due to other viruses or diseases other than Hepatitis C (e.g. Hepatitis B, Alcoholics....etc.) were excluded from the study.

Patients and controls were subjected to the followings:

- Full history taking and complete clinical examination.
- Laboratory investigations: Complete blood count, liver biochemical profile, viral markers.
- Radiological investigations for patients include: Abdominal ultrasonography, triphasic CT abdomen and/or dynamic MRI abdomen with MRI diffusion.
- The presence of the viral genome in serum was assessed by using the Amplicor reverse-transcription PCR assay (Roche, Welwyn, UK).
- The patients of the second group were subjected to liver biopsy to determine degree of fibrosis.
- *Apolipoprotein E* Genotyping for all the samples obtained from patients and control group was done by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method.

Methodology

ApoE genotyping was performed by extracting DNA from whole blood samples using DNA extraction kit supplied from Qiagen (Valencia, CA, and USA). Genotyping for *ApoE* polymorphisms was carried out as method described by (Hixson and Vernier, 1990). Amplification reactions were performed, DNA was amplified in a 50 µL reaction mixture using PCR master mix supplied by Qiagen. Thermo cycling was done using in a thermal cycler (Perkin-Elmer 9600, Cambridge, United Kingdom) according to the following cycle profile: initial denaturation for 2 minutes at 95°C, 40 cycles at denaturation 95°C (1 min.), annealing 60°C (1 min.), extension 72°C (2 min.) and for the final phase, there was an initial incubation at 72°C (5 min.). The primers were used for amplification by PCR, with the following sequences (Emi et al., 1988): Forward 5'- TAAGCTTGGCACGGCTGTCCAAGGA-3', Reverse 5'-ACAGAATTCGCCCGGCTGGTACAC-3'. Upon completion of PCR, the products were analyzed by electrophoresis on a 2% ethidium bromide-stained agarose gel. After PCR amplification of *ApoE* region, 5 units of *HhaI* (New England Biolabs). *HhaI* cleaves at GCGC encoding 112arg (ε4) and 158arg (ε3, ε4), but does not cut at GTGC encoding 112cys (ε2, ε3) and 158cys (ε2) (Table 1). *HhaI* was added directly to each reaction mixture for digestions of *ApoE* sequences for >3

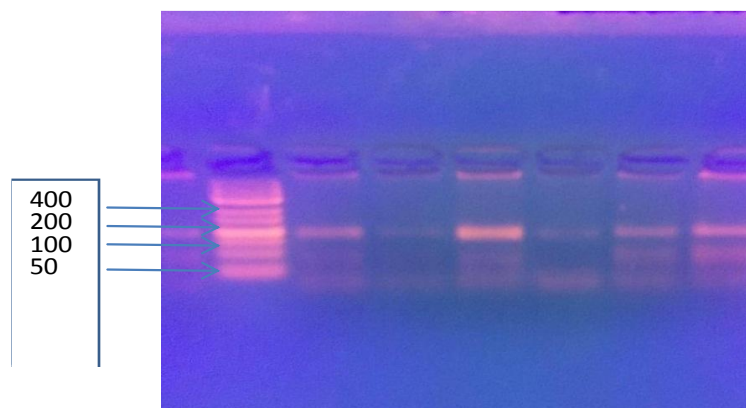


Figure 1. Electrophoretic separation of *HhaI* fragments after gene amplification of DNA from subjects with known *ApoE* isoforms. The $\epsilon 3/\epsilon 3$ samples contained the 91 bp fragment (112cys), as well as 48 fragment, the 35 bp fragments (not shown). The fragment sizes (in bp) of a DNA standard (Thermo Scientific Gene Ruler Low Range DNA ladder) are shown to the left of the gel. The $\epsilon 3/\epsilon 4$ samples not shown.....

hours at 37°C; blocking of the reaction was done at 65°C for 15 minutes. Each reaction mixture was loaded onto 8% Polyacrylamide Gel (1.5 mm thick and 25 cm long), and electrophoresed for 3 hours under constant current (45 mA). After electrophoresis, the gel was treated with ethidium bromide (0.2 mg/dl) for 10 minutes, and DNA fragments were visualized by UV illumination. The sizes of *HhaI* fragments were estimated by comparison with known size markers (Thermo Scientific GeneRuler Low Range DNA ladder).

Statistical analysis

The collected data was organized, tabulated and statistically analyzed using statistical package for social science (SPSS) computer package, version 17 (SPSS Inc, USA), running on IBM-compatible computer. Quantitative data were represented as mean, standard deviation (SD).

Qualitative (categorical) data were represented as relative frequency and percent distribution, Chi square (X^2) for comparison between groups was used. Allele frequencies are reported with their group percentages. The risk associated with the presence of the *ApoE*-4 allele was estimated for each diagnostic subgroup by using odds ratios (with their 95% confidence limits). For interpretation of results, p value ≤ 0.05 was considered statistically significant.

RESULTS

Baseline characteristics of CHC patients are demonstrated in (Table 1). Mean age was 45.85 ± 5.26 in CHC (non cirrhotic) patients, 54.30 ± 10.38 in CHC (cirrhotics/LF) and 52.50 ± 10.57 in healthy controls

without significant difference between groups ($p > 0.05$). Non significant difference regarding sex between the studied groups. The study groups were also different regarding symptoms (hepatic encephalopathy and ascites), and imaging studies (ultrasonography for liver and spleen) with highly significant statistical difference ($p < 0.001$).

The laboratory data among studied patients

On comparing the study groups as regards laboratory investigations, it was found that there was highly significant difference in TLC, Hb and Plt counts ($p < 0.001$ for all parameters). As regards liver function tests, a significant difference in ALT, AST ($p = 0.03, 0.04$), and a highly significant difference in total bilirubin and serum albumin ($p < 0.001$ for both parameters). As regards coagulation profile, there was highly significant difference between the study groups ($p < 0.001$ for all parameters).

Restriction enzymes *HhaI* digestion for genotype assays for *APOE* Gene

PCR amplification shows 244 bp fragments, each genotype was determined by fragments combination specific for each genotype after restriction enzymes *HhaI* digestion. $\epsilon 3/\epsilon 3$ sample contained the 91 bp fragment (112cys) and due arginine at amino acid position 158 in $\epsilon 3$, the 158 fragment was digested to 48 and 35 bp fragments in $\epsilon 3$ allele, in addition, $\epsilon 4/\epsilon 4$ has arginine at position 112, so 91 bp fragment will be digested to unique 72 bp and 19 bp (short to be detected). 38 bp, and (16bp and 18bp; short to be detected) (common fragment for all genotypes) (Figure 1).

Table 2. Distribution of the genotypes and alleles frequencies of *ApoE* in the studied groups

Genotypes	CHC (Non cirrhotic)		CHC (Cirrhotic/LF)		Controls		Chi-square	
	n	%	n	%	n	%	X ²	P-value
($\epsilon 2/2$ - 3 - 4)/($\epsilon 4/ \epsilon 4$)	0	0.00	0	0.00	0	0.00	2.83	0.24
($\epsilon 3/ \epsilon 3$)	35	87.50	39	97.50	36	90.00		
($\epsilon 3/ \epsilon 4$)	5	12.50	1	2.50	4	10.00		
Total	40	100.00	40	100.00	40	100.00		
Allele frequency	N	N	N	N	N			
$\epsilon 2$	0	0.00	0	0.00	0	0.00	2.71	0.25
$\epsilon 3$	75	93.75	79	98.75	76	95.00		
$\epsilon 4$	5	6.25	1	1.25	4	5.00		
Total	80	100.00	80	100.00	80	100.00		

Note: n, number of subjects; N, number of alleles.

Table 3. The association between *ApoE* Gene Polymorphism and HCV progression to liver cirrhosis

Genotypes	CHC	
	LC/LF vs Non cirrhotic	P
($\epsilon 2/2$ - 3 - 4)/($\epsilon 4/ \epsilon 4$)	-	-
($\epsilon 3/ \epsilon 3$)	Reference	
($\epsilon 3/ \epsilon 4$)	0.18 (0.02 to 1.61)	0.12
Allele frequency	OR (CI95%)	
$\epsilon 2$	-	-
$\epsilon 3$	Reference	
$\epsilon 4$	0.19 (0.02 to 1.66)	0.13

The genotypes and alleles frequency of *APOE* Gene Polymorphism

Among the 120 subjects, the most common genotype was $\epsilon 3/ \epsilon 3$, accounting for (91.67)%, followed by $\epsilon 3/ \epsilon 4$ (8.33) %. The genotypes of $\epsilon 2/2$ $2/3$ $2/4$ and $\epsilon 4/4$ were not detected in our results. The study reported that; *ApoE* ($\epsilon 3/ \epsilon 3$) wild homozygous genotype (97.50)% (87.50)% (90) % in CHC with LC with signs of liver cell failure (LF), non cirrhotic patients and healthy control and *ApoE* heterozygous genotype ($\epsilon 3/ \epsilon 4$) (2.50)% (12.50)% (10.00) % in those groups respectively. The $\epsilon 3$ allele was the most common allele over represented in CHC LC/LF (98.75) % versus (93.75)% in non cirrhotic group. However, CHC non cirrhotic patients had a higher *ApoE* $\epsilon 4$ allele frequency (6.25) % than those with severe disease (1.25) % (Table 2). Major contribution of *ApoE* $\epsilon 4$ allele was with decreased susceptibility to LC/LF development can not be ruled out; (OR) (CI95), 0.18(0.02 to 1.61)/0.19 (0.02 to 1.66) ($p=0.12,0.13$) ($\epsilon 3/ \epsilon 4$) genotype and $\epsilon 4$ allele frequency respectively (Table 3).

DISCUSSION

Hepatitis C virus infection is a major global health prob-

lem. HCV infection affects almost 3% of the world's population. More than 170 million people worldwide are infected with HCV (Shepard et al., 2005; Kamal and Nasser, 2008) with the highest prevalence in Egypt (15%) with genotype 4 being the most common (Kamal and Nasser, 2008; World Health Organization, 2007; El-Zanaty and Way, 2009). The consequences of chronic infection also vary (Wozniak et al., 2002); some individuals develop minor or no liver damage, whereas others suffer from progressive chronic hepatitis, leading to liver cirrhosis, and even HCC (Alter et al., 1992; Frank et al., 2000; Bostan and Mahmood, 2010; Wozniak et al., 2016) and high rates of infection are observed among persons in all age groups (Abdel-Aziz et al., 2000; Perz et al., 2006), the reasons for the diversity of outcomes of HCV infection are unclear, infection with a particular genotype is not thought to influence disease outcome. Alternatively, host factors may play a role; some of these factors include male sex, an older age at infection, increased alcohol intake, coinfection with HIV-1 or HBV (Simmonds et al., 1993; Zarski et al., 1998; Mohsen, 2001) and HLA types (Khakoo et al., 2004), genetic factors could be one of major influence.

ApoE is multifunctional protein with numerous roles in lipoprotein metabolism. The frequencies of the *apoE*

Table 4. Estimated worldwide distribution of human *ApoE* allele frequencies

<i>Estimated worldwide human allele frequencies of ApoE based upon over 200 world populations and 50,000 people (highly variable depending upon population)</i>			
<i>Allele</i>	$\epsilon 2$	$\epsilon 3$	$\epsilon 4$
<i>Frequency</i>	0 – 37.5%	8.5 – 98%	0 – 49%

alleles vary between different ethnicities. Several global studies on *ApoE* genotype frequency varied greatly among different population that *ApoE* ($\epsilon 3/\epsilon 3$) genotype, the most common genotype in different population followed by ($\epsilon 4/\epsilon 4$) and then ($\epsilon 2/\epsilon 2$) (Hallman et al., 1991). The $\epsilon 3$ allele is usually the most prevalent, present in 50-90% of individuals, whereas $\epsilon 2$ has the lowest frequency at 0-15%, and is even absent in some native populations. The $\epsilon 4$ allelic variant occurs at a frequency of 5-30% (Gerdes et al., 1996). Also, Farrer et al., (1997) stated that the estimated worldwide distribution of *ApoE* $\epsilon 4$ allele was 13.7%. Another study by Eisenberg et al., (2010) Table 4.

Previous studies on *ApoE* genotypes revealed gene polymorphism and its role in susceptibility to different diseases and many infectious agents; Hill et al., (2007) stated possible role of *ApoE* $\epsilon 2$ in HCV viral clearance via defective binding of HCV lipoviral particle to cellular receptors involve in entry of this virus particle. In further contrast, *ApoE* $\epsilon 2$ is a risk factor for herpes simplex encephalitis (Wozniak et al., 2002). Whereas *ApoE* $\epsilon 4$ it is a risk factor for Alzheimer's disease in those harboring HSV1 in the brain, and a risk factor for cold sores in those harboring HSV1 in the peripheral nervous system (Itzhaki et al., 1997; Lin et al., 1998). Also, it confers a risk for reversible dementia and peripheral neuropathy in HIV-infected people (Corder et al., 1998).

ApoE $\epsilon 4$ is considered as an accepted risk factor for Alzheimer's disease and cardiovascular disorders. Recently, associations between *APOE* $\epsilon 4$ and lower probability of progression to HCV-related liver cirrhosis has been demonstrated. The possible mechanism for *ApoE* $\epsilon 4$ action, the amino acid arginine substitution results in an "interaction between *ApoE* $\epsilon 4$ amino- and carboxyl- terminal domains, known as "domain interaction" (Mahley and Huang, 2009), this structural difference is believed to play a role in the altered function of *ApoE* $\epsilon 4$ " (Villeneuve et al., 2014), another explanation stated by Davignon et al., (1988), *ApoE* $\epsilon 4$ allele is associated with increased serum levels of LDL, HCV entry through LDLRs, (Monazahian et al., 1999). The greater number of LDLR ligands in the serum of *ApoE* $\epsilon 4$ allele carrier lead to competition for the receptor binding, therapy may prevent or reduce binding of the virus to hepatocytes and therefore reduce cell damage. In this study we investigated whether specific *ApoE* genotypes and HCV induced liver cirrhosis in Egyptian patients by PCR-RFLP. PCR amplification and Restriction isotyping methodology avoids the use of costly and time-

consuming hybridization and sequencing techniques (Hixson and Vernier, 1990).

The study reported that *ApoE* heterozygous genotype ($\epsilon 3/\epsilon 4$) and $\epsilon 4$ allele more frequent in CHC patients (non cirrhotic) than decompensated group with decreased susceptibility to LC/LF development can not be excluded. Although non statistically significant difference due to relatively small sample size, with a relatively larger sample size, it may achieve statistical significance confirmed this figure. Compared to the study of Wozniak et al. (2002) in which *ApoE* $\epsilon 4$ allele frequency was 6.5% in cirrhotic patients vs. 20% in non-cirrhotic patients, our results is similar to Wozniak's study in the concept that the cirrhotic group had a lower *ApoE* $\epsilon 4$ allele frequency than the non-cirrhotic group, which further proves the hypothesis that *ApoE* $\epsilon 4$ genotype may have a role in protection against HCV-induced liver cirrhosis. Another study by Mueller et al., (2016) reported lower-representation of *ApoE* $\epsilon 4$ allele carriers in patients with CHC compared to the higher-representation of *APOE* $\epsilon 4$ allele carriers in patients who spontaneously cleared HCV infection suggests a protective role of the *ApoE* $\epsilon 4$ allele in the course of HCV infection.

The estimated worldwide distribution of *ApoE* $\epsilon 4$ allele widely varied frequency, the prevalence of *ApoE* $\epsilon 4$ allele frequency in our study 5.00%, this difference can be attributed to possible ethnic factors related to Egyptian population, or small sample.

CONCLUSION

The effect of *ApoE* on disease risk is complex. More studies still warranted to understand; the factors and mechanisms involved in *ApoE*-related interactions which could improve awareness of disease susceptibility, disease progression, disease prevention, and *APO E* status and therapeutic association. Assessing *ApoE* genotypes may give valuable information that help to guide medical decisions in clinical practice and for personalized medicine. Limitation of the current study, this study involving only 80 CHC Egyptian patients and 40 healthy controls, for this relatively small sample size and estimated worldwide overall prevalence of *ApoE* $\epsilon 4$ allele frequency and the difference related to Egyptian population, probability and statistics may have been a limiting factor in the detection of the statistically significant association between *ApoE* gene polymorphism in CHC patients. Our result support many different other

studies and highlight possible role of *ApoE* ϵ 4 Allele for protection with lower probability of progression to HCV-related liver cirrhosis in Egyptian patients but further investigations with a planned prospective population based with different ethnicity with larger sample size are needed.

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Conflict of Interest

The authors declare no conflict of interest

REFERENCES

- Abdel-Aziz F, Habib M, Mohamed MK, Abdel-Hamid M, Gamil F, Madkour S, Mikhail NN, Thomas D, Fix AD, Strickland GT, Anwar W, Sallam I (2000). Hepatitis C virus (HCV) infection in a community in the Nile Delta: population description and HCV prevalence. *Hepatology*; 32(1): 111-5.
- Agnello V, Abel G, Elfahal M, Knight GB, Zhang QX (1999). Hepatitis C virus and other flaviviridae viruses enter cells via low density lipoprotein receptor. *Proc Natl Acad Sci U S A*; 96(22):12766-71.
- Ahn SJ, Kim DK, Kim SS, Bae CB, Cho HJ, Kim HG, Kim YJ, Lee JH, Lee HJ, Lee MY, Kim KB, Cho JH, Cho SW, Cheong JY (2012). Association between apolipoprotein E genotype, chronic liver disease, and hepatitis B virus. *Clin Mol Hepatol*; 18(3): 295-301.
- Aizawa Y, Seki N, Nagano T, and Abe H (2015). Chronic hepatitis C virus infection and lipoprotein metabolism. *World J Gastroenterol*; 21(36): 10299-313.
- Alter MJ, Margolis HS, Krawczynski K, Judson FN, Mares A, Alexander WJ, Hu PY, Miller JK, Gerber MA, Sampliner RE, et al. (1992). The natural history of community-acquired hepatitis C in the United States. *N Engl J Med*; 327 (27):1899-905.
- Bennet AM, Di Angelantonio E, Ye Z, Wensley F, Dahlin A, Ahlborn A, Keavney B, Collins R, Wiman B, de Faire U, Danesh J (2007). Association of apolipoprotein E genotypes with lipid levels and coronary risk. *JAMA*; 298(11):1300-11.
- Bostan N, Mahmood T (2010). An overview about hepatitis C: a devastating virus. *Crit Rev Microbiol*; 36:91-133.
- Bowen DG and Walker CM (2005). Adaptive immune responses in acute and chronic hepatitis C virus infection. *Nature*; 436 (7053): 946-52.
- Corder EH, Robertson K, Lannfelt L, Bogdanovic N, Eggertsen G, Wilkins J, Hall C (1998). HIV-infected subjects with the E4 allele for APOE have excess dementia and peripheral neuropathy. *Nat Med*; 4(10):1182-4.
- Das HK, McPherson J, Bruns GAP, Karathanasis SK, Breslow JL (1985). Isolation, characterization, and mapping to chromosome 19 of the human apolipoprotein E gene. *J Biol. Chem.*; 260: 6240-7.
- Davignon J, Gregg RE, Sing CF (1988). Apolipoprotein E polymorphism and atherosclerosis. *Arteriosclerosis*. 8(1):1-21.
- Eichner JE, Dunn ST, Perveen G, Thompson DM, Stewart KE, Stroehla BC (2002). Apolipoprotein E polymorphism and cardiovascular disease: a HuGE review. *Am J Epidemiol*; 155(6):487-95.
- Eisenberg DTA, Kuzawa CW, Hayes MG (2010). Worldwide allele frequencies of the human apolipoprotein E (APOE) gene: climate, local adaptations and evolutionary history. *Ame. J. Physical Anthropol.*; 143 (1): 100–111.
- El-Zanaty F, Way A (2009). *Egypt Demographic and Health Survey 2008*, Egyptian: Ministry of Health (El-Zanaty and Associates and Macro International, Cairo):1-431.
- Emi M, Wu LL, Robertson MA, Myers RL, Hegele RA, Williams RR, White R, Lalouel JM (1988). Genotyping and sequence analysis of apolipoprotein E isoforms. *Genomics*; 3(4): 373-9.
- Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, Myers RH, Pericak-Vance MA, Risch N, Van Duijn CM (1997). Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta-Analysis Consortium. *JAMA*; 278(16):1349-56.
- Frank C, Mohamed MK, Strickland GT, Lavanchy D, Arthur RR, Magder LS, El Khoby T, Abdel-Wahab Y, AlyOhn ES, Anwar W, Sallam I (2000). The role of parenteral antischistosomal therapy in the spread of hepatitis C virus in Egypt. *Lancet*; 355(9207):887-91.
- Gerdes LU, Gerdes C, Hansen PS, Klausen IC, Faergeman O, Dyerberg J (1996). The apolipoprotein E polymorphism in Greenland Inuit in its global perspective. *Hum Genet*, 98(5):546- 50.
- Hallman DM, Boerwinkle E, Saha N, Sandholzer C, Menzel HJ, Császár A, Utermann G (1991). The apolipoprotein E polymorphism A: Comparison allele frequencies and effects in nine population. *Am. J. Hum. Genet.* 49(2): 338-49.
- Hill JM, Bhattacharjee PS, Neumann DM (2007). Apolipoprotein E alleles can contribute to pathogenesis of numerous clinical conditions including HSV1-corneal disease. *Exp Eye Res*; 84(5):801-11.
- Hishiki T, Shimizu Y, Tobita R, Sugiyama K, Ogawa K, Funami K, Ohsaki Y, Fujimoto T, Takaku H, Wakita T, Baumert T F, Miyanari Y, Shimotohno K (2010). Infectivity of hepatitis C virus is influenced by association with apolipoprotein E isoforms. *J. Virol.*; 84(22): 12048-57.
- Hixson JE and Vernier DT (1990). Restriction isotyping of human apolipoprotein E by gene amplification and cleavage with HhaI. *J. Lipid Res.*; 31: 545-8.
- Itzhaki RF, Lin WR, Shang D, Wilcock GK, Faragher B, Jamieson GA (1997). Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. *Lancet*; 349 (9047):241-4.
- Kamal SM and Nasser IA (2008). Hepatitis C Genotype 4: What we know and what we don't yet know. *Hepatology*; 47(4):1371-83.
- Kelly ME, Clay MA, Mistry MJ, Hsieh-Li HM, Harmony JA (1994). Apolipoprotein E inhibition of proliferation of mitogen-activated T lymphocytes: production of interleukin 2 with reduced biological activity. *Cell Immunol*; 159(2):124-39.
- Khakoo SI, Thio CL, Martin MP, Brooks CR, Gao X, Astemborski J, Cheng J, Goedert JJ, Vlahov D, Hilgartner M, Cox S, Little AM, Alexander GJ, Cramp ME, O'Brien SJ, Rosenberg WM, Thomas DL, Carrington M (2004). HLA and NK cell inhibitory receptor genes in resolving hepatitis C virus infection. *Science*; 305(5685):872–4.
- Lin WR, Graham J, MacGowan SM, Wilcock GK, Itzhaki RF (1998). Alzheimer's disease, herpes virus in brain, apolipoprotein E4 and herpes labialis. *Alzheimers Rep*; 1:173-8.
- Mahley RW, Huang Y (2009). Alzheimer disease: multiple causes, multiple effects of apolipoprotein E4 and multiple therapeutic approaches. *Ann. Neurol.*; 65(6): 623-5.
- Minihane AM, Jofre-Monseny L, Olano-Martin E, Rimbach G (2007). ApoE genotype, cardiovascular risk and responsiveness to dietary fat manipulation. *Proc Nutr Soc*; 66:183-97.
- Mohsen AH (2001). The Trent HCV Study Group. The epidemiology of hepatitis C in a UK health regional population of 5.12 million. *Gut*; 48(5):707-13.
- Monazahian M, Bohme I, Bonk S, Koch A, Scholz C, Grethe S, Thomssen R (1999). Low density lipoprotein receptor as a candidate receptor for hepatitis C virus. *J Med Virol*; 57(3): 223-9.
- Mueller T, Fischer J, Gessner R, Rosendahl J, Böhm S, Florian van Bommel, Knop V, Sarrazin C, Witt H, Marques AM, Kovacs P, Schleinitz D, Stumvoll M, Bluher M, Bugert P, Schott E and Berg T (2016). Apolipoprotein E allele frequencies in chronic and self-limited hepatitis C suggest a protective effect of APOE4 in the course of hepatitis C virus infection. *Liver International*.
- Myklebost O, Rogne S (1988). A physical map of the apolipoprotein gene cluster on human chromosome 19. *Hum Genet.* 78(3): 244-7.

- Nielsen SU, Bassendine MF, Burt AD, Martin C, Pumeechockchai W, Toms GL (2006). Association between hepatitis C virus and very-low-density lipoprotein (VLDL)/LDL analyzed in iodixanol density gradients. *J. Virol.* 2006; 80(5): 2418-28.
- Ordovas JM, Litwack-Klein L, Wilson PW, Schaefer MM, Schaefer EJ (1987). Apolipoprotein E isoform phenotyping methodology and population frequency with identification of apoE1 and apoE5 isoforms. *J Lipid Res*; 28(4):371-80.
- Perz JF, Armstrong GL, Farrington LA, Hutin YJ, Bell BP (2006). The contributions of hepatitis B virus and hepatitis C virus infections to cirrhosis and primary liver cancer worldwide. *J Hepatol*; 45(4): 529-38.
- Poynard T, Bedossa P, Opolon P (1997). Natural history of liver fibrosis progression in patients with chronic hepatitis C. The OBSVIRC, METAVIR, CLINIVIR, and DOSVIRC groups. *Lancet*, 349 (9055): 825-32.
- Sabile A, Perlemuter G, Bono F, Kohara K, Demaugre F, Kohara M, Matsuura Y, Miyamura T, Bréchet C, Barba G (1999). Hepatitis C virus core protein binds to apolipoprotein AII and its secretion is modulated by fibrates. *Hepatology*; 30:1064-76.
- Sacre SM, Stannard AK, Owen JS (2003). Apolipoprotein E (ApoE) isoforms differentially induce nitric oxide production in endothelial cells. *FEBS Lett*;540(1-3):181-7.
- Shepard CW, Finelli L, Fiore AE, Bell BP (2005). Epidemiology of hepatitis B and hepatitis B virus infection in United States children. *Pediatr Infect Dis J*; 24(9): 755-60.
- Shore VG, and Shore B (1973). Heterogeneity of human plasma very low density lipoproteins. Separation of species differing protein components. *Biochemistry*;12: 502-7.
- Simmonds P, Holmes EC, Cha TA, Chan SW, McOmish F, Irvine B, Beall E, Yap PL, Kolberg J, Urdea MS (1993). Classification of hepatitis C virus into six major genotypes and a series of subtypes by phylogenetic analysis of the NS-5 region. *J Gen Virol*;74 (Pt 11):2391-9.
- Villeneuve S, Brisson D, Marchant NL and Gaudet D (2014). The potential applications of Apolipoprotein E in personalized medicine *Front Aging Neurosci*;6:154.
- Weisgraber KH, Rall SC Jr, Mahley RW (1981). Human E apoprotein heterogeneity. Cysteine-arginine interchanges in the amino acid sequence of the apo-E isoforms. *J. Biol. Chem*; 256(17):9077-83.
- World Health Organization (2007). Hepatitis C Available: <http://www.who.int/mediacentre/factsheets/fs164/en>
- Wozniak MA, Lugo Iparraguirre LM, Dirks M2, Deb-Chatterji M, Pflugrad H, Goldbecker A, Tryc AB, Worthmann H, Gess M, Crossey MM4, Forton DM, Taylor-Robinson SD, Itzhaki RF (2016). Apolipoprotein E deficiency and cognitive function in hepatitis C virus infected patients. *Journal of viral hepatitis*; 23: 39-46.
- Wozniak MA, Itzhaki RF, Faragher EB, James MW, Ryder SD, Irving WL (2002). Apolipoprotein E- epsilon 4 Protects against severe liver disease caused by hepatitis C virus. *Hepatology*; 36(2): 456-63.
- Zannis VI, and Breslow JL (1981). Human very low density lipoprotein apolipoprotein E isoform polymorphism is explained by genetic variation and post-translational modification. *Biochemistry.*; 20(4): 1033-41.
- Zarski JP, Bohn B, Bastie A, Pawlotsky JM, Baud M, Bost-Bezeaux F, Tran van Nhieu J, Seigneurin JM, Buffet C, Dhumeaux D (1998). Characteristics of patients with dual infection by hepatitis B and C viruses. *J Hepatol*;28(1):27-33.